Structure–Immunosuppressive Activity Relationships of New Analogues of 15-Deoxyspergualin. 1. Structural Modifications of the Hydroxyglycine Moiety

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A series of new analogues of 15-deoxyspergualin (DSG), an immunosuppressive agent currently commercialized in Japan, was synthesized and tested in a graft-versus-host disease (GVHD) model in mice. Using the general concept of bioisosteric replacement, variations of the hydroxyglycine central "C" region were made in order to determine its optimum structure in terms of in vivo immunosuppressive activity. By this way, the malonic derivative **13a** was discovered as the first example of a new series of potent immunosuppressive agents encompassing a retro-amide bond linked to the hexyl-guanidino moiety. Structure–activity relationships of this series were studied by synthesizing compounds **13g–i** and **13k–s**. Variation of the "right-amide" of **13a** led to the urea **19a** and the carbamates **23** and **27a** which proved to be equally active as DSG in our GVHD model. Finally **27a** was found to be the most potent derivative, being slightly more active than DSG in a heart allotransplantation model in rats. Due to the absence of chiral center in its structure and to its improved chemical stability compared to DSG, **27a** was selected as a candidate for clinical evaluation.

Introduction

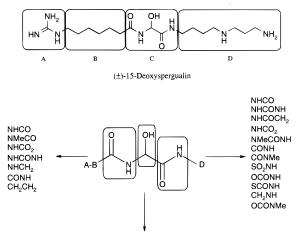
In the last three decades, major advances have been achieved in the development of immunosuppressive drugs which have made possible organ transplantation and treatment of autoimmune diseases. Azathioprine¹ was first introduced to control graft rejection and is still used in combination with other immunosuppressive agents. However, its myelotoxicity and other side effects limit its use. The discovery of cyclosporin A² allowed major improvements in transplantation. It induced dramatic increase of organ allograft survival which made possible liver and cardiac transplants. More recently, FK-506³ has been introduced as a more potent immunosuppressant although mechanistically related to cyclosporin A. These two immunosuppressive compounds block T-cell activation via the inhibition of interleukin-2 expression.⁴ Recently, mycophenolate mofetil⁵ which inhibits the purine biosynthesis has been approved for the prevention of kidney graft rejection. However, although more specific than azathioprine, these drugs do show nephro-, neuro-, and myelotoxicities as well as other side effects.⁶ Therefore, there is a need for new less toxic compounds and with new mechanisms of action, which could lead to a more specific immunosuppression.

Among the different compounds recently developed in this area (-)15-deoxyspergualin $((-)DSG)^7$ appeared to be attractive considering its biological profile and its new mechanism of action.⁸ (-)DSG is a derivative of (-)spergualin,⁹ a fermentation product isolated from *Bacillus laterosporus* by Umezawa's group. It was first obtained by dehydroxylation of (-)spergualin in 1982.¹⁰ Its first total synthesis was published in 1987,¹¹ and despite its low overall yield,¹² this process afforded (-)-DSG and (+)DSG which could be separately evaluated. In vivo experiments clearly showed that (–)DSG is the only immunosuppressive enantiomer.⁷ However, most of the biological, pharmacological, and clinical data have been obtained with the more readily available (+,–)-DSG whose first total synthesis was patented in 1982.¹³ Since its discovery, DSG has been widely studied and demonstrated efficacy first as an anticancer agent,¹⁴ then in animal transplantation either in allografts¹⁵ or xenografts,¹⁶ and more recently in graft-versus-host disease (GVHD) animal models.¹⁷ DSG has also been widely investigated¹⁸ in autoimmune diseases. The high potential of DSG to reverse renal graft rejection was first reported in 1990.¹⁹ This drug was therefore launched in Japan (Spanidin) in this indication in 1994.¹⁹

However DSG suffers from three main drawbacks which limit its use. The first one is related to its low chemical stability in aqueous solution where hydrolysis of the hydroxyglycine part leads to 7-guanidinoheptanamide and glyoxyloylspermidine. This cleavage is a problem for large scale synthesis, purification, and standard galenic development. The second problem is due to the presence of the inactive (+)DSG isomer present in the racemic mixture. This compound is not reported to be immunosuppressive but can contribute to the acute toxicity.⁷ Until now, industrial synthesis of the active (-)DSG form or another equivalent achiral compound was not available. The last limitation is related to a very low oral bioavailability (less than 5%²⁰), preventing oral administration.

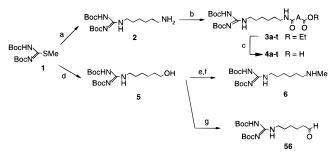
We present here the first part of our program aimed at the discovery of new DSG analogues (Chart 1), devoid of the above-mentioned drawbacks. For this the DSG structure has been divided into four regions. The purpose of the present study was to find the optimal "C" region which could alleviate problems associated with chirality and hydrolytic sensitivity brought by the hydroxyglycine moiety. Umezawa and co-workers have

[†] Deceased.



 $\begin{array}{l} \mathsf{CHOH},\ \mathsf{CHOCH}_3,\ \mathsf{CHF},\ \mathsf{CHNH}_2,\ \mathsf{CHNHAc},\ \mathsf{CHOCH}_2\mathsf{Ph},\ \mathsf{C}(\mathsf{CCH}_3)_{2^1},\ \mathsf{C}(\mathsf{CH}_2\mathsf{OH})_2,\\ \mathsf{CHCH}_3,\ \mathsf{CHCH}_2\mathsf{CH}_3,\ \mathsf{CHPh},\ \mathsf{CHBn},\ \mathsf{C}(\mathsf{CH}_3)_2,\ \mathsf{(CH}_2)_{n\ =\ 0,1,2,3},\ \mathsf{CH=CH}(\mathsf{E},\mathsf{Z}),\ \mathsf{CH}_2\mathsf{CH}(\mathsf{OH}),\\ \mathsf{(CH}_2)_2\mathsf{NH},\ \mathsf{(CH}_2)_2\mathsf{O}. \end{array}$

Scheme 1^a



^{*a*} Reagents: (a) hexanediamine, THF; (b) HOOCACOOEt, CH_2Cl_2 , DCC, HOBT; (c) 1 M NaOH, DME (v/v); (d) amino-6-hexanol, THF; (e) Et_3N , CH_2Cl_2 , -20 °C, methanesulfonyl chloride; (f) DME, MeNH₂ (40%); (g) PDC, CH_2Cl_2 .

already explored the structure–activity relationship (SAR) of this part of the molecule, synthesizing numerous analogues where the hydroxyglycine has been replaced by various α -, β -, and γ -amino acids.²¹ *O*methylhydroxyglycine, L-serine, and glycine were the only tolerated α -amino acids. γ -Aminobutyric and γ -amino α - or β -hydroxybutyric acids were also tolerated, but none of these derivatives were as active as DSG. β -Amino acids and all other α -amino acids led to inactive compounds. This stringent structural requirement for this "C" region is remarkable. However, all these analogues retained the two amide bonds with the same orientation as in the parent DSG. We thus decided to extend the investigation of this crucial part of the

Scheme 2^a

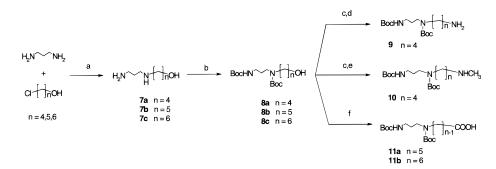
molecule using concepts encountered in peptide medicinal chemistry such as retro-inverso approach and other bioisosteric replacements for the amide bonds. We also replaced the hydroxymethylene moiety by various linkers (Chart 1).

Chemistry

The wide range of structural modifications planned forced us to use rather diverse synthetic schemes. Moreover, the choice of these schemes was dictated by availability of certain starting materials.

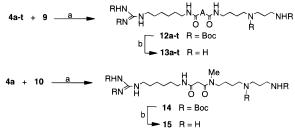
The malonamide-related compounds 13a-t and 15 were made by the route illustrated in Schemes 1, 2, and 3 from key intermediates **4a**-**t**, **9**, and **10**. Compounds **4a**-**t** were prepared in three steps from *N*.*N*-bis(*tert*butoxycarbonyl)-S-methylisothiourea 1 which was obtained according to the procedure described by Bergeron et al.²² (Scheme 1). Condensation of 1 with hexanediamine led to the intermediate **2** which was coupled to commercially available monoester malonic acid derivatives using standard coupling techniques. Saponification of the ester groups led to compounds 4a-t. We used a shorter procedure (Scheme 2) than the one described by Bergeron et al.²³ to obtain 9. Alkylation of propane diamine by *n*-chloroalkanol in refluxing butanol led to compounds 7a-c with good yields. The amines were then protected by Boc groups to give **8a-c**. Conversion of alkanol 8a to its methanesulfonate derivative followed by nucleophilic displacement with ammonia or methylamine yielded the desired spermidine derivatives 9 and 10. Target compounds 13a-t and 15 were obtained by the coupling of key intermediates **4a**-**t** with **9** or **4a** with 10 using standard coupling techniques followed by removal of Boc protecting groups of the intermediates **12a**-t or **14** with trifluoroacetic acid (TFA) in CH₂Cl₂ (Scheme 3).

The synthetic sequences utilized for the preparation of derivatives 19a-c, 21a-b, 23, 35, and 42 are outlined in the Scheme 4. The key intermediates 17a-cwere obtained by coupling 2 with *N*-benzyloxycarbonylglycine, *N*-benzyloxycarbonyl-*N*-methyl-glycine, or *N*-benzyloxycarbonyl- β -alanine using standard coupling techniques followed by cleavage of the Cbz protecting group with Pd/C under a hydrogen atmosphere. The synthesis of urea derivatives (19a-c) was best achieved using a strategy involving condensation of 17a-c with bis(4-nitrophenyl)carbonate followed by the action of 9 to give the N-Boc protected compounds 18a-c. Final



^{*a*} Reagents: (a) KI, K₂CO₃, n-BuOH reflux; (b) (Boc)₂O, dioxane, H₂O; (c) Et₃N, CH₂Cl₂, -20 °C, methanesulfonyl chloride; (d) EtOH, NH₄OH (v/v); (e) EtOH, MeNH₂; (f) PDC, DMF.

Scheme 3^a





deprotection was then performed to provide the desired DSG analogues **19a**-**c**. In the same way, we adopted a similar strategy for the synthesis of the carbamate **23**. To the nonisolated intermediate nitrophenyl carbonate, generated by reaction of 4-nitrophenyl chloroformate with **8a**, was added **17a** to afford **22**, which was further deprotected to yield the desired carbamate derivative **23**. Oxidation of alkanol **8b**-**c** by pyridinium dichromate in DMF provided the carboxylic derivatives **11a**-**b** (Scheme 2). Compounds **21a**-**b** were obtained via the coupling of **17a** with intermediates **11a**-**b** to yield **20a**-**b**. Cleavage of the Boc protecting groups was then carried out by the standard method, i.e., TFA in CH₂-Cl₂ to give the target compounds **21a**-**b**.

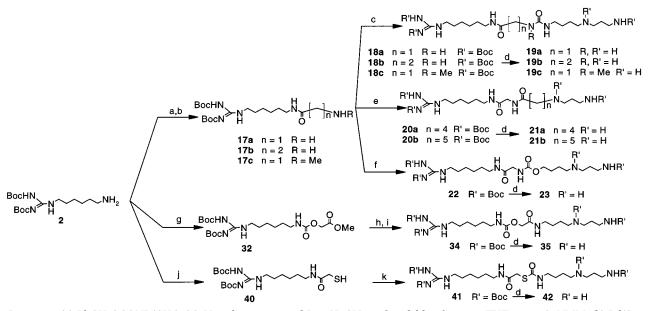
Addition of **2** to the methyl [(phenoxycarbonyl)oxy] acetate generated from methyl glycolate and phenyl

Scheme 4^a

chloroformate led to the intermediate **32**. Saponification of **32** with NaOH in DME followed by coupling with **9**, according to the standard techniques, and deprotection afforded **35**. Compound **42** was prepared via the key intermediate **40** which was obtained by the coupling of **2** with thioglycolic acid. Compound **40** was further converted to the thiocarbamate **42** by reaction of the nonisolated nitrophenyl thiocarbonate with **9** and deprotection with the standard acidic procedure.

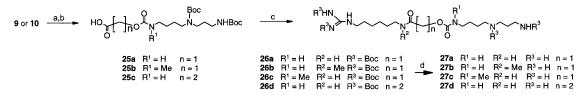
The synthetic route leading to the DSG analogues **27a**-**d**, where the carbamate function is reversed compared to **23**, is outlined in Scheme 5. Acylation of spermidine derivatives **9** or **10** (Scheme 2) with phenyl carbonates derived from methyl hydroxyacetate or hydroxypropionate afforded, after saponification, **25a**-**c**. These acids were converted to the final products **27a**-**d** by coupling with **2** or **6** (Scheme 1) followed by removal of protecting groups.

The compounds **31**, **39**, **46**, **50**, **55**, and **60** were made starting from **9** using specific routes illustrated in Scheme 6 and which are explained below. The intermediate 10-[(phenylmethoxy)carbonylamino]decanoic acid was coupled with **9** using standard coupling technique to give **28**. Removal of the Cbz group by hydrogenolysis, condensation with **1**, and final deprotection yielded the desired compound **31**. Carbobenzyloxyglycine was coupled to **9**, and the Cbz group of the coupling product **36** was then cleaved. The obtained intermediate was then



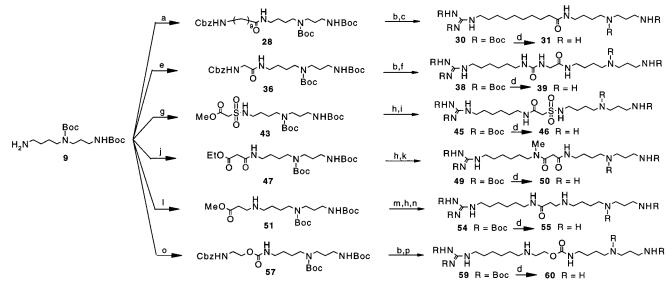
^{*a*} Reagents: (a) PhCH₂OCONR(CH₂)_{*n*}CO₂H with n = 1, 2 and R = H, CH₃, isobutylchloroformate, THF, -20 °C, NMM; (b) Pd/C 10%, EtOH, H₂; (c) bis-(4-nitrophenyl)carbonate, THF, **9**; (d) CH₂Cl₂, TFA (v/v); (e) **11a** or **11b**, isobutylchloroformate, THF, -30 °C, NMM; (f) **8a**, Et₃N, THF, 4-nitrophenyl chloroformate; (g) methyl[(phenoxycarbonyl)oxy]acetate, toluene, 40 °C; (h) 1 M NaOH, DME (v/v); (i) DCC, HOBT, CH₂Cl₂, **9**; (j) thioglycolic acid, carbonyldiimidazole, CH₂Cl₂; (k) 4-nitrophenyl chloroformate, Et₃N, THF then **9**.

Scheme 5^a



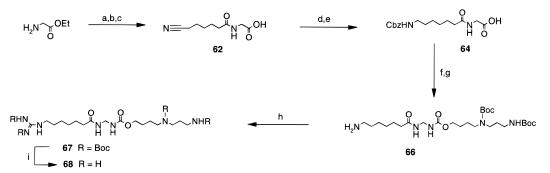
^{*a*} Reagents: (a) PhOCO₂(CH₂)_{*n*}CO₂Me with n = 1, 2, toluene, reflux; (b) 1 M NaOH, DME (v/v); (c) DCC, HOBT, CH₂Cl₂, **2** or **6**; (d) CH₂Cl₂, TFA (v/v).

Scheme 6^a



^{*a*} Reagents: (a) CbzHN(CH₂)₉CO₂H, CHCl₃, DCC, HOBT; (b) Pd/C 10%, EtOH, H₂; (c) **1**, THF; (d) CH₂Cl₂, TFA (v/v); (e) carbobenzyloxyglycine, CHCl₃, DCC, HOBT; (f) 4-nitrophenyl chloroformate, THF, Et₃N, **2**; (g) methyl 2-(chlorosulfonyl)acetate, CH₂Cl₂, -20 °C, Et₃N; (h) 1 M NaOH, DME (v/v); (i) carbonyldiimidazole, CH₂Cl₂, Et₃N, **2**; (j) ethyl hydrogen malonate, CHCl₃, DCC, HOBT; (k) CH₂Cl₂, DCC, HOBT, **6**; (l) methylacrylate, MeOH reflux; (m) (Boc)₂O, Et₃N, THF; (n) isobutyl chloroformate, THF, -30 °C, NMM, **2**; (o) PhOCO₂(CH₂)₂NHCbz, Et₃N, toluene; (p) EtOH, **56**, AcOH then NaBH₃CN.

Scheme 7^a



^{*a*} Reagents: (a) 6-bromohexanoic acid, CHCl₃, DCC, HOBT; (b) EtOH, reflux, KCN; (c) 1 M NaOH, DME (v/v); (d) H₂, Ni Raney, EtOH, 1 M NaOH; (e) EtOH, NaHCO₃, benzylchloroformate; (f) Et₃N, THF, DPPA then **8a**, toluene, reflux; (g) H₂, Pd/C 5%, EtOH; (h) THF, **1**; (i) CH₂Cl₂, TFA (v/v).

reacted with 4-nitrophenylchloroformate to give a reactive carbamate derivative which in the presence of **2** at room temperature provided the desired protected compound. Final Boc protecting groups cleavage was then performed to give **39**.

Methyl methyl 2-(chlorosulfonyl)acetate reacted with **9** in dichloromethane to afford the key intermediate **43**. Then usual sequence involving saponification, amide bond coupling of **2** using carbonyldiimidazole, and final deprotection was performed to obtain **46**.

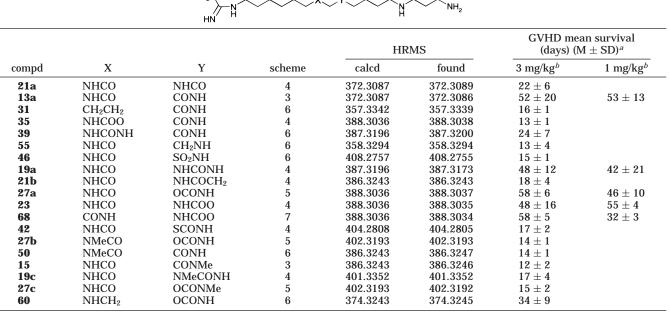
The reactions carried out to obtain **50** are similar to those used for **46**; however, they include coupling **6** instead of **2** and using the DCC, HOBT procedure.

Michael addition of **9** to methylacrylate in refluxing methanol afforded the key intermediate **51** in which the amine was further protected by a Boc group. Saponification, coupling with **2** using the mixed anhydride method, and final deprotection allowed us to obtain the desired compound **55**. *N*-carbobenzyloxy-2-aminoethanol reacted with phenyl chloroformate in toluene to afford the reactive carbonate which in the presence of **9** led to the intermediate **57**. Removal of Cbz group of **57** by hydrogenolysis afforded the corresponding amine which was converted to the final compound **60** by a reductive amination reaction with **56** (see Scheme 1) in the presence of NaBH₃CN followed by cleavage of Boc protecting groups.

The synthetic sequence leading to **68** with a reversed amide bond compared to 23 is outlined in Scheme 7. N-(6-bromohexanoyl)-glycine ethyl ester was obtained by coupling bromohexanoic acid with glycine ethyl ester. Nucleophilic substitution of the bromine by KCN followed by saponification of the ester allowed the production of the acid **62**. Hydrogenation of the nitrile group carried out with Raney nickel in the presence of sodium hydroxide afforded the amine which was protected by a Cbz group to give 64. The crucial conversion of 64 to 66 was carried out as follows: reaction of 64 with diphenylphosphorazide gave the corresponding acyl azide derivative which upon Curtius rearrangement in toluene at reflux led to the corresponding isocyanate which was further reacted with 8a (Scheme 2) to give 66 after cleavage of the Cbz protecting group. Reaction of the primary amine of 66 with 1 (Scheme 1) in THF and further acidic cleavage of the Boc protecting groups afforded the desired diacylaminal compound 68.

Table 1. Physicochemical and Biological Data of 15-DSG Analogues with X and Y Variations

 H_2N



^{*a*} The mean survival of control is 15 ± 2 days and, respectively, 38 ± 12 and 57 ± 7 days for animals treated with DSG at 1 mg/kg and 3 mg/kg. ^{*b*} All compounds were administered daily by i.p. route.

Results and Discussion

Biological and analytical data are shown in Tables 1, 2, and 3. Immunosuppressive activity was assayed in vivo in a mouse GVHD model which was adapted from the literature.²⁴ Briefly, GVHD is induced at day 0, treatments are then given from day 1 to day 10 (day 6 omitted), and survival is followed until sacrifice at day 60. The mean survival of the controls is 15 ± 2 days. In such conditions, DSG administered by i.p. route increases survival to 38 ± 12 days at 1 mg/kg/day and to 57 ± 7 days at 3 mg/kg/day. All compounds were then studied by the same route first at 3 mg/kg/day and then at 1 mg/kg/day if they demonstrated a significant activity. The first step of our investigation dealt with the modification of the two amide bonds. Our goal was to get more stable compounds that were easier to synthesize and devoid of chirality. Thus, we decided to make glycine instead of hydroxyglycine analogues of DSG. We first synthesized 21a (Table 1) where the two amide bonds are reversed compared to DSG. This modification which destroys the spermidine integrity completely abolishes the immunosuppressive activity.

However **13a**, where only the "left amide" was reversed compared to DSG, was active at 3 and 1 mg/kg i.p. Other replacements of the "left amide" such as a two-methylene unit (**31**), a carbamate (**35**), or an urea (**39**) gave inactive compounds. Similarly, bioisosteric replacement of the "right amide" with a reduced amide (**55**) or a sulfonamide (**46**) was detrimental to activity. By contrast, **19a**, which can be seen as an analogue of inactive compound **21a** where the nitrogen of spermidine has been reintroduced giving a urea, was active at 3 mg/kg i.p.. Further modification of the urea of **19a** was successful and afforded potent and less toxic analogues.²⁵ Either NH can be favorably replaced by an O atom leading to **27a** and **23** which were very active at 3 and 1 mg/kg i.p. and which were in the same range

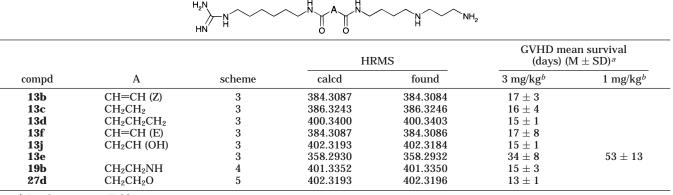
of activity as that of DSG. However, NH cannot be replaced by a methylene (**21b**). It is worth noting that compound **23** was the first example of a very active analogue of DSG not encompassing the full spermidine moiety. The carbamate is clearly a very suitable function in this position whatever the orientation of the "left amide" (see activity of **68**). Interestingly the corresponding thiocarbamate (**42**) was inactive, possibly due to its poor stability at physiological pH. N-Methylation of either the amides **27b**, **50**, and **15**, the urea **19c**, or the carbamate **27c** was absolutely detrimental. Finally, reduction of the "left amide" of **27a** gave rise to the slightly active compound **60**.

Table 2 shows results obtained with analogues of **13a**, **19a**, and **27a** where the nature and length of the central linker have been modified. Obviously a single bond (**13e**) or a methylene (**13a**) were the only tolerated linkers. It is interesting to notice that a CH_2CH_2 linker leads to a totally inactive compound (**13c**) when its replacement with a CH_2O (**27a**) affords one of the most active derivatives of this series (Table 1).

In Umezawa's derivatives, the presence of OH or, to a lesser extent, of OCH₃ in the "C" region increases the potency compared to hydrogen. We investigated (Table 3) the effect of such groups or related ones such as F, NH₂, or NHAc on compound **13a**. We also introduced lipophilic aliphatic and aromatic substituents. Finally we made some disubstituted derivatives. In sharp contrast to the DSG series, introduction of OH (**13s**) or OCH₃ (**13i**) does not really increase activity. However, only OH, OCH₃, or F (**13t**) groups are tolerated in contrast to NH₂ (**13p**), NHAc (**13o**), or OCH₂Ph (**13r**) which, though susceptible to sharing a hydrogen bond, are detrimental for activity.

Disubstitution with OCH₃ (13q) and CH₂OH (13l) is also totally ineffective. Finally, as it is the case for DSG series, alkyl or aryl substituents led to inactive compounds 13g, 13h, 13k, 13m, and 13n.

Table 2. Physicochemical and Biological Data of 15-DSG Analogues with A Modifications



a,b See footnotes in Table 1.

 Table 3. Physicochemical and Biological Data of Malonic Derivatives with R1 and R2 Substituents

			HRMS		GVHD mean survival (days) $(M \pm SD)^a$	
compd	R_1	R_2	calcd	found	3 mg/kg ^b	1 mg/kg ^l
13s	OH	Н	388.3036	388.3030	53 ± 13	12 ± 1
13i	OCH_3	Н	402.3193	402.3193	55 ± 12	27 ± 3
13t	F	Н	390.2993	390.2996	60 ± 0	18 ± 11
13p	NH_2	Н	387.3196	387.3196	16 ± 1	
130	NHAc	Н	429.3302	429.3299	16 ± 1	
13r	OCH ₂ C ₆ H ₅	Н	478.3506	478.3501	14 ± 2	
13q	OCH ₃	OCH_3	432.3298	432.3298	12 ± 3	
131	CH ₂ OH	CH ₂ OH	432.3298	432.3296	17 ± 2	
13h	CH_3	Н	386.3243	386.3243	16 ± 2	
13m	CH_2CH_3	Н	400.3400	400.3399	19 ± 3	
13k	C_6H_5	Н	448.3400	448.3403	19 ± 4	
13g	CH ₂ C ₆ H ₅	Н	462.3556	462.3553	16 ± 2	
13n	CH ₃	CH_3	400.3400	400.3407	15 ± 1	

^{*a,b*} See footnotes in Table 1.

Table 4. Prevention of Graft Rejection after Heart Allotransplantation in the Dark Agouti to Lewis Rat Combination

compd	dose ^a (mg/kg)	survival (day)	mean survival (day) (M \pm SD)	statistical analysis ^b
control	0	6, 6, 6, 7, 7, 7, 7	7 ± 1	
DSG	6	13, 20, 20, 26, 42, 46, >100	38 ± 30	S
27a	6	17, 27, 27, 45, 63, 99, >100	54 ± 34	S
13a	6	5, 7, 8, 10, 10, 13	9 ± 3	NS
13e	6	11, 11, 11, 11, 11, 11, 15	12 ± 2	S
19a	6	6, 6, 12, 12, 13, 15, 17	12 ± 4	S
13s	6	15, 20, 24, 25, 28, 37, 79	33 ± 22	S

^{*a*} All compounds were administered daily by i.p. route for 10 days starting the day after surgery. ^{*b*} Manntel and Haenszel test according to SAS (Cary, NC).

Selection of the best immunosuppressor compounds obtained in this study was then made using a more demanding model. Some of the more active compounds were then tested in a model for acute organ rejection after heart allotransplantation in the rat. Genetic combination was selected to be completely histoincompatible, and the recipient was considered to be a high responder in which control of rejection would be especially difficult.^{26,27} DSG induces a 38 ± 30 days survival of the graft in these experimental conditions (10 days, 6 mg/kg, i.p.) whereas rejection is observed at 7 ± 1 days in controls (Table 4). Despite its good activity in the mouse GVHD test, 13a was unable to control graft rejection in rats at 6 mg/kg. Compounds 13e and 19a were only slightly active in this model, showing that it is a good discrimination step. However **13s**, which can

be seen as a retro-analogue of DSG, induced a 4- to 5-fold graft survival increase compared to the control. Finally, 27a proved to be the best tested compound of the series, being slightly more active than DSG. This compound was thus selected as a candidate for further preclinical investigation. In particular, it proved to be far more stable than DSG as assessed by its chemical stability studies. Figure 1 shows the kinetics of degradation of DSG and 27a in aqueous solution at different pH at 37 °C. DSG chemical stability is clearly pH dependent. Its half-life is around 15 days at pH 1, 2 days at pH 7, and a few hours at pH 10. As anticipated, the stability of 27a is much better: at acidic or neutral pH, 27a degradation is less than 10% after 50 days. However at pH 10, its half-life is around 1 day. This high improvement in chemical stability makes the handling

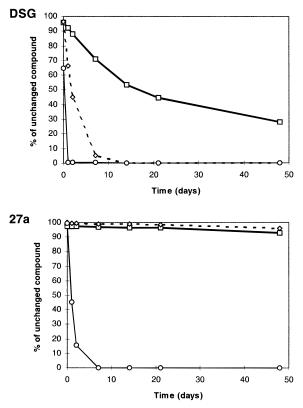


Figure 1. The chemical stability of DSG and **27a** in aqueous buffers at pH 1 (\Box), pH 7 (\diamond), and pH 10 (\bigcirc) at 37 °C.

of **27a** much easier in aqueous medium. In particular, purification, formulation, and dosing can be carried out without any problem of degradation. This practical aspect of this freely water soluble compound intended to be administered by injection is crucial for its development.

Conclusion

In the present study we have synthesized a series of new DSG analogues in order to investigate the structural requirement for the so-called "C" region of the molecule in term of immunosuppressive activity. It proved to be rather stringent. However, we have shown that the left amide bond can be present in both directions but other functions such as urea, carbamate, and a two methylene unit are not compatible. N-Methylation of this amide is also precluded. Nevertheless the right amide bond can be favorably replaced by a carbamate in both directions and less successfully with an urea. N-Methylation of these functionalities led to inactive compounds. The central linker should be either a single bond or a methylene which can also be monosubstituted by a methoxy or hydroxyl group or by a fluorine atom.

This study allowed us to select some DSG analogues which have the same range of potency as DSG without containing chirality. Moreover, their chemical stability was strongly improved in comparison to DSG, which makes their industrial synthesis much more accessible. Due to its optimum biological profile in terms of immunosuppressive properties in the heart allograft model^{28,29} and in a GVHD model after bone marrow transplantation³⁰ and also in terms of safety pharmacology and toxicology, **27a** was selected as a drug candidate and is currently undergoing clinical evaluation.

Experimental Section

Chemistry. ¹H Nuclear magnetic resonance spectra were obtained on a Bruker 300 at 300 MHz. The chemical shifts are expressed in δ values (part per million) relative to tetramethylsilane as internal standard. Significant ¹H NMR data are reported in the following order: multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet), number of protons.

The identity of final compounds was verified by exact mass spectral determinations performed by the University of Rennes I (Centre Régional de Mesures Physiques de l'Ouest). Highresolution mass spectra (HRMS) and low resolution mass spectra (LRMS) were performed using a ZABSpec TOF from V. G. Analytical (L-SIMS ionization mode, Cs⁺, mNBA).

Thin-layer chromatography was performed on silica gel $60F_{254}$ plates or RP-18F_{254S} (Merck). Merck silica gel 60 (230–400 mesh) was used for flash column chromatography and lichroprep RP-18 Merck (5–20 μ m) for medium-pressure liquid chromatography.

DSG was prepared following Umezawa's procedure.14

General Procedure for the Preparation of Compounds 13a-t and 15. Compounds of the general structure **13a-t** (Scheme 3) were synthesized from carboxylic acid derivatives **4a-t**, and 4-aminobutyl[3-[(1,1-dimethylethoxy)carbonylamino]propyl]carbamic acid, 1,1-dimethyl-ethyl ester **9** by the representative procedure illustrated for analogue **13a**.

Carboxylic acid derivatives 4a-t were obtained in three steps (Scheme 1) from *N*,*N*-bis(*tert*-butoxycarbonyl)-*S*-methoxy isothiourea 1 which was prepared according to the procedure described by R. J. Bergeron and J. S. MacManis.²³

The compounds **9** and **10** were prepared in three steps as follows (Scheme 2).

4-[(3-Aminopropyl)amino]butanol (**7a**). 4-Chloro-1-butanol (2.2 g, 20.0 mmol) was added into a solution of 1,3diaminopropane (3.0 g, 40.0 mmol), potassium iodide (0.33 g, 2.0 mmol), and potassium carbonate (1.4 g, 10.0 mmol) in butanol (50 mL). The mixture was stirred at reflux for 24 h. Then the mixture was slowly cooled to room temperature, filtered, and concentrated under vacuum to afford **7a** as a colorless oil which was used without further purification (2.8 g, 86% yield). ¹H NMR (DMSO-*d*₆) δ : 1.4–1.5 (m, 6H), 2.4– 2.6 (m, 6H), 3.4–3.5 (m, 2H).

(4-Hydroxybutyl)[3-(1,1-dimethylethoxy)carbonylamino]propyl]carbamic Acid, 1,1-Dimethylethyl Ester (8a). This compound was obtained from 4-[(3-aminopropyl)amino]butan-1-ol (2 g, 13.67 mmol) according to the procedure described by F. Veznik et al.³¹ as a colorless oil (4.45 g, 94% yield).

(4-Aminobutyl)[3-[(1,1-dimethylethoxy)carbonylamino]propyl]carbamic Acid, 1,1-Dimethylethyl Ester (9). To a stirred solution of **8a** (1 g, 2.9 mmol) and triethylamine (0.7 g, 3.76 mmol) in CH_2Cl_2 (10 mL) cooled to -20 °C was added dropwise a solution of methanesulfonyl chloride (0.43 g, 3.76 mmol) in CH_2Cl_2 (5 mL). The stirring was continued at -20 °C for 10 min, and then the reaction mixture was allowed to warm to room temperature. An additional portion of CH_2Cl_2 was added, and the solution was washed with saturated K_2CO_3 solution. The layers were separated, and the aqueous phase was extracted with CH_2Cl_2 . The organic layers were combined, dried over MgSO₄, filtered, and concentrated under vacuum.

The obtained crude product was taken up in ethanol (20 mL) and ammonium hydroxide (20 mL). The reaction mixture was then stirred for 48 h at room temperature. The solvents were removed under vacuum, and the oily residue was purified by flash chromatography on silica gel (EtOAc then EtOAc/EtOH/ NH₄OH 6/3/0.1) to afford **9** as a clear oil (0.46 g, 46%). ¹H NMR (CDCl₃) δ : 1.4–1.6 (m, 24H), 2.7 (t, 2H), 3.1–3.25 (m, 6H), 4.8 (s, 1H), 5.3 (s, 1H).

[4-(Methylamino)butyl] [3-[(1,1-dimethylethoxy)carbonyl amino]propyl]carbamic Acid, 1,1-Dimethylethyl Ester (10). Starting with 8a, the same procedure described for the synthesis of 9 but using methylamine as nucleophilic agent was applied to 10 obtained in 72% yield. ¹H NMR (CDCl₃) δ : 1.4–1.8 (m, 24H), 2.7 (s, 3H), 2.9–3.3 (m, 8H), 4.8– 5.4 (br s, 1H), 8.1–9.5 (br s, 1H).

5-[*N*-[3](1,1-dimethylethoxy)carbonylamino]-propyl]-*N*-[(dimethylethoxy)carbonyl]-amino]-pentanoic Acid (11a). Pyridinium dichromate (77.5 g, 206.0 mmol) was added in small portions to a solution of **8b** (21.2 g, 58.88 mmol) in DMF (100 mL). The reaction medium was stirred for 6 h at room temperature, then diluted with H₂O (700 mL) and extracted with Et₂O (3×150 mL). The organic phase was washed with a CuSO₄ solution, dried, concentrated, and purified by flash chromatography on silica gel (EtOAc/hexane 5/5 then EtOAc) to afford **11a** (20.0 g, 91% yield) as a yellow oil. ¹H NMR (CDCl₃) δ : 1.4–1.7 (m, 24H), 2.3 (t, 2H), 3.0–3.3 (m, 6H), 4.7 (br s, 1H), 5.3 (br s, 1H).

[(6-Aminohexyl)carbonimidoyl]bis-carbamic Acid, Bis-(1,1-dimethylethyl) Ester (2). Hexane-1,6-diamine (17.23 g, 0.148 mol) was added at room temperature to a stirred solution of *N*,*N*-bis (*tert*-butoxycarbonyl)-*S*-methylisothiourea (43.0 g, 0.148 mol) in THF (300 mL). The reaction medium was stirred for 16 h. After evaporation of the solvent, the obtained residue was purified by flash chromatography on silica gel (CHCl₃/ EtOH 3/1 then EtOAc/MeOH/32% aqueous ammonia mixture 6/3/0.1) to afford **2** (19.7 g, 37%) as a yellow oil. ¹H NMR (CDCl₃) δ : 1.25–1.60 (m, 28H), 2.7 (t, 2H), 3.5 (q, 2H), 8.3 (t, 1H), 11.5 (s, 1H).

3-[[(1,1-Dimethylethoxy)carbonyl]amino]-12-oxo-2,4,-11-triaza-tetradec-2-enedioic Acid, 1-[1,1-Dimethylethyl Ester]14-ethyl Ester (3a). To a stirred solution of ethyl hydrogen malonate (3.7 g, 28.0 mmol) in CH₂Cl₂ (50 mL) were added at 0 °C 1,3-dicyclohexyl carbodiimide (DCC) (5.8 g, 28.0 mmol) and 1-hydroxybenzotriazole hydrate (HOBT) (0.4 g, 3 mmol). The mixture was allowed to stand for half an hour at 0 °C. Then 2 (5 g, 14 mmol) in solution in CH₂Cl₂ (30 mL) was added dropwise at 0 °C. After the addition was complete, the mixture was allowed to warm gradually to room temperature overnight. The solvent was evaporated off under reduced pressure, and the oily residue was purified by flash chromatography on silica gel (EtOAc/methylcyclohexane 1/1) to afford **3a** (6.27 g, 95% yield) as a viscous oil. ¹H NMR (CDCl₃) δ : 1.2– 1.7 (m, 29H), 3.2-3.3 (m, 4H), 3.4 (q, 2H), 4.2 (q, 2H), 7.2 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

3-[[(1,1-Dimethylethoxy)carbonyl]amino]-12-oxo-2,4,-11-triaza-tetradec-2-enedioic Acid, 1-[1,1-Dimethylethyl Ester] (4a). 3a (1.97 g, 4.19 mmol) was dissolved in a mixture of 1 N NaOH (20 mL) and DME (20 mL). The reaction mixture was stirred for 1 h at room temperature, concentrated to a third of its volume, and then acidified to pH 2 with 1 N HCl. Extraction of the water layer with 2 × 50 mL of CHCl₃, followed by evaporation of the organic phases under reduced pressure, afforded a residue which was purified by flash chromatography on silica gel (EtOAc/EtOH 1/9) to give **4a** (1.29 g, 70%). ¹H NMR (CDCl₃) δ : 1.2–1.7 (m, 24H), 3.1–3.4 (m, 6H), 7.7 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

3-[[(1,1-Dimethylethoxy)carbonyl]amino]-20-[(1,1-dimethylethoxy)carbonyl]-12,14-dioxo-2,4,11,15,20,24-hexaazapentacosenedioic Acid, Bis(1,1-dimethylethyl) Ester (12a). To a stirred solution of 4a (1.25 g, 2.8 mmol) in CH₂Cl₂ (20 mL) were added at 0 °C DCC (0.57 g, 2.8 mmol) and HOBT (0.07 g, 0.5 mmol). The mixture was allowed to stand for 0.5 h at 0 °C. Then 9 (1 g, 2.9 mmol) in solution in CH₂Cl₂ (10 mL) was added dropwise at 0 °C. After the addition was complete, the mixture was allowed to warm gradually to room temperature and left to stand one night. The solvent was evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (EtOAc/methylcyclohexane 1/1 then EtOAc/MeOH 9/1) to afford **12a** (1.6 g, 73%) as a viscous oil. ¹H NMR (CDCl₃) δ : 1.3-1.7 (m, 50H), 3.1-3.3 (m, 12H), 3.4 (q, 2H), 4.8 and 5.3 (br s, 1H), 6.8 and 7.15 (br s, 2H), 8.3 (t, 1H), 11.5 (s, 1H).

N-[4-[[3-(Amino)propyl]amino]butyl]-*N*-[6-[(aminoiminomethyl)amino]hexyl]propanediamide, Tris(trifluoroacetate) (13a). 12a (0.7 g, 0.9 mmol) was dissolved in 10 mL of TFA and 10 mL of anhydrous CH₂Cl₂. The reaction mixture was stirred for 24 h at room temperature, and the solvents were then removed under reduced pressure. The obtained residue was taken up in 150 mL of distilled water and then lyophilized. The residue was purified by MPLC (medium-pressure liquid chromatography) on a RP18 silica (type grafted, particle size 5 to 20 μ m) using a water/CH₃CN/TFA mixture (7/2/1, v/v/v) as eluent to afford **13a** (0.43 g, 66% yield) in the form of a translucent white amorphous solid. ¹H NMR (DMSO-*d*₆) δ : 1.2–1.6 (m, 12H), 1.9 (m, 2H), 2.9–3.1 (m, 14H), 7.2 (br s, 4H), (t, 1H), 8 (m, 5H), 8.7 (br s, 2H). ¹³C NMR (DMSO-*d*₆) δ : 22.5, 23.4, 25.4, 25.6, 25.7, 28.0, 28.5, 35.8, 37.6, 38.2, 40.3, 43.0, 43.5, 46.1, 156.6, 166.9 (2C).

3-[[(1,1-Dimethylethoxy)carbonyl]amino]-20-[(1,1-dimethylethoxy)carbonyl]-15-methyl-12,14-dioxo-2,4,11,-15,20,24-hexaazapentacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (14). 14 (1,15 g, 65%) was obtained in the form of an oil according to the method described for 12a starting from 4a (1 g, 2.25 mmol) and 10 (0.86 g, 2.4 mmol). ¹H NMR (CDCl₃) δ : 1.3–1.7 (m, 50H), 3.1–3.3 (m, 15H), 3.4 (q, 2H), 4.8 and 5.3 (br s, 1H), 6.8 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

N-[4-[(3-Aminopropyl)amino]butyl]-*N*-[6[(aminoiminomethyl)amino]hexyl]-*N*-methyl-propanediamide, Tris-(trifluoroacetate) (15). 15 (0.72 g, 72%) was obtained as an oil according to the method described for 13a starting from 14 (1 g, 1.27 mmol). ¹H NMR (DMSO- d_6) δ : 1.2–1.6 (m, 12H), 1.8–2.0 (m, 2H), 2.8–3.1 (m, 13H), 3.2–3.4 (m, 4H), 7.8–8.0 (m, 3H), 8.0–8.1 (m, 1H), 8.5–8.8 (m, 4H). ¹³C NMR (DMSO- d_6) δ : 23.5, 24.1, 24.5, 26.2, 26.3, 26.4, 28.1, 28.9, 35.2, 36.5, 37.3, 40.3, 41.9, 45.2, 48.2, 158.5, 168.6, 168.8.

General Procedure for the Preparation of Compounds 27a–d. Compounds of the general structure **27a–d** (Scheme 5) were synthesized from carboxylic acid derivatives **25a–d** and **2** or **6** by the representative procedure illustrated for **27a**. Carboxylic acid derivatives **25a–d** were obtained in two steps from **9** or **10**. **6** was prepared in three steps as follows (Scheme 1).

Preparation of Compound 6. [(6-Hydroxyhexyl)carbonimidoyl]biscarbamic Acid, Bis(1,1-dimethylethyl) Ester (5). Amino-6 hexanol (4.24 g, 35.0 mmol) was added at room temperature, with stirring, to a solution of N,N'-bis (*tert*-butoxycarbonyl)-S-methylisothiourea (9,54 g, 32.8 mmol) in THF (100 mL). The reaction medium was stirred for 4 days. After evaporation of the solvent, the obtained residue was purified by flash chromatography on silica gel (methylcyclohexane/EtOAc 7.5/2.5) to afford 5 (9.05 g, 77% yield). ¹H NMR (CDCl₃) δ : 1.2–1.7 (m, 26H), 3.35–3.5 (m, 2H), 3.6–3.7 (m, 2H), 8.2–8.4 (br s, 1H), 11.5 (br s, 1H).

[(6-Aminomethylhexyl)carbonimidoyl]biscarbamic Acid, Bis(1,1-dimethylethyl) Ester (6). To a stirred solution of 5 (2 g, 5.57 mmol) and triethylamine (1.4 g, 13.8 mmol) in CH_2Cl_2 (20 mL) cooled to - 20 °C was added dropwise a solution of methanesulfonyl chloride (0.8 g, 7 mmol) in CH₂- Cl_2 (5 mL). The stirring was continued at -20 °C for 15 min, and the mixture was then allowed to warm to room temperature. An additional portion of CH₂Cl₂ was added, and the solution was washed with a saturated K₂CO₃ solution. The layers were separated, and the aqueous phase was extracted with CH₂Cl₂. The organic layers were combined, dried over MgSO₄, filtered, and concentrated under vacuum. The obtained crude product was taken up in DME (5 mL) and methylamine in aqueous solution (40%) (6.2 mL, 80 mmol). The reaction mixture was then stirred for 18 h. The solvents were removed under vacuum, and the oily residue was taken up in EtOAc (100 mL) and washed with a brine solution. The organic layer was dried over MgSO₄, filtered, and concentrated under vacuum. The obtained oily residue was purified by flash chromatography on silica gel (EtOAc/EtOH/NH4OH 6/3/0.1) to afford **6** as a yellow oil (1.7 g, 83% yield). ¹H NMR (CDCl₃) δ : 1.3-1.7 (m, 26H), 2.50 (s, 3H); 2.68 (t, 2H), 3.40 (t, 2H), 8.3 (t, 1H), 11.5 (s, 1H).

Preparation of Compound 27a. 6-[(1,1 Dimethylethoxy)carbonyl]-13-oxa-12-oxo-2,6,11-triazapentadecanedioic Acid, 1-(1,1-Dimethylethyl) Ester 15-Methyl Ester (24a). A solution of **9** (7.4 g, 21.4 mmol) in toluene (20 mL) was added to a stirred solution of methyl [(phenoxycarbonyl) oxy] acetate (4.5 g, 21.4 mmol) in toluene (100 mL). The reaction medium was refluxed for 15 h. The solvent was evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (methylcyclohexane/EtOAc 7/3 and then EtOAc) to afford **24a** (8.4 g, 85%) as an oil. ¹H NMR (CDCl₃) δ : 1.45–1.65 (m, 24H), 3.05–3.25 (m, 8H), 3.8 (s, 3H), 4.7 (s, 2H), 4.9 and 5.3 (br s, 1H).

6[(1,1-Dimethylethoxy)carbonyl]-13-oxa-12-oxo-2,6,11triazapentadecanedioic Acid, 1-(1,1-Dimethylethyl Ester) (25a). This compound was obtained (6.7 g, 81% yield) as an oil by following a procedure analogous to preparation of **4a** and starting from **24a** (8.45 g, 18.3 mmol). ¹H NMR (CDCl₃) δ : 1.3–1.8 (m, 24H), 3.10–3.25 (m, 8H), 4.7 (s, 2H), 5.0 (t, 1H), 6.8 (s, 1H).

3-[[(1,1-Dimethylethoxy)carbonyl]amino]-21-[(1,1-dimethylethoxy)carbonyl]-14-oxa-12,15-dioxo 2,4,11,16,21,-25-hexaazahexacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (26a). This compound was obtained (3.85 g, 73% yield) in the form of an oil by following a procedure analogous to preparation of 12a and starting from 25a (3.0 g, 6.7 mmol) and 2 (2.4 g, 6.7 mmol). ¹H NMR (CDCl₃) δ : 1.35–1.65 (m, 50H), 3.10–3.30 (m, 12H), 4.55 (s, 2H), 5.2–5.5 (br s, 1H), 6.4 (br s, 1H).

4-[3-(Aminopropyl)amino]butylcarbamic Acid, 2-[[6-[(aminoiminomethyl)amino]-hexyl]amino]-2-oxoethyl Ester, Tris(trifluoroacetate) (27a). This compound was obtained as a translucent white amorphous solid (2.61 g, 73% yield) according to the procedure described for **13a** and starting from **26a** (3.85 g, 4.9 mmol). ¹H NMR (DMSO-*d*₆) δ : 1.25–1.55 (m, 12H), 1.9 (m, 2H), 2.80–3.10 (m, 12H), 4.35 (s, 2H), 5.5– 8.6 (m, 12H). ¹³C NMR (D₂O) δ : 23.6, 24.5, 26.2, 26.3, 26.7, 28.6, 29.0, 37.3, 39.8, 40.6, 41.9, 45.2, 48.2, 63.7, 157.5, 158.2, 171.4.

General Procedure for the Preparation of Compounds 19a-c, 21a-b, and 23. Compounds of the general structure 19a-c, 21a-b, and 23 were synthesized according to the Scheme 4 by the representative procedures illustrated for analogues 19a, 21a, and 23.

Preparation of Compound 19a. 3-[[(1,1-Dimethylethoxy)carbonyl]amino]-12-oxo-2,4,11,14-tetraazapentadec-2enedioic Acid, 1-(1,1-Dimethylethyl) Ester 15-Phenylmethyl Ester (16a). A solution of isobutyl chloroformate (1.6 g, 14.0 mmol) in THF (5 mL) was added dropwise to a solution cooled to -30 °C of carbobenzyloxyglycine (3.0 g, 14.0 mmol) and N-methylmorpholine (2.8 g, 28 mmol) in THF (50 mL). The reaction medium was stirred for 0.5 h and a solution of 2 (5.4 g, 14 mmol) in THF (20 mL) was added. Stirring was maintained for 2 h at -30 °C and then for 24 h at room temperature. After filtration of the reaction medium and evaporation of the filtrate under vacuum, the residue obtained was purified by flash chromatography on silica gel (EtOAc/ methylcyclohexane 1/1) to afford 16a as an oil (7.16 g, 91% yield). ¹H NMR (CDCl₃) δ: 1.3-1.7 (m, 26H), 3.2 (q, 2H), 3.4 (q, 2H), 3.8 (d, 2H), 5.15 (s, 2H), 5.5 (br s, 1H), 6.0 (br s, 1H), 7.3 (s, 5H).

13-Amino-3-[[(1,1-dimethylethoxy)carbonyl]amino] 12-oxo-2,4,11-triazatridec-2-enoic Acid, 1,1-Dimethylethyl Ester (17a). A mixture of **16a** (7.1 g, 13 mmol) and 10% palladium on carbon (0.7 g) in ethanol (120 mL) was stirred at room temperature and under a hydrogen atmosphere for 2 h at atmospheric pressure. The catalyst was then filtered off, and the organic phase was evaporated to give **17a** (5.3 g, 98% yield) in the form of an oily residue which was used without further purification for the preparation of **18a**. ¹H NMR (CDCl₃) δ : 1.3–1.6 (m, 28H), 3.25–3.45 (m, 6H), 7.3 (s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

3-[[(1,1-Dimethylethoxy)carbonyl]amino]-21-[(1,1-dimethylethoxy)carbonyl]-12,15-dioxo-2,4,11,14,16,21,25heptaazahexacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (18a). Bis(4-nitrophenyl)carbonate (4.3 g, 13 mmol) was added in small portions to a solution of **17a** (5.3 g, 12 mmol) in anhydrous THF (50 mL). The reaction medium was stirred for 1 h at room temperature, and a solution of **9** (4.5 g, 13 mmol) in anhydrous THF (50 mL) was added dropwise. Stirring was continued for 24 h at room temperature, and the solvent was evaporated off under reduced pressure. The obtained residue was purified by flash chromatography on silica gel (EtOAc) to afford **18a** (6.01 g, 64% yield) as an oil. ¹H NMR (CDCl₃) δ : 1.3–1.7 (m, 50H), 3.1–3.35 (m, 12H), 3.8 (d, 2H), 4.8 and 5.8 (br s, 3H), 6.9 (t, 1H), 8.3 (t, 1H); 11.5 (s, 1H).

N-[4-[[3-(Amino)propyl]amino]butyl]-*N*-[[[[6-[(aminoiminomethyl)amino]hexyl]amino]carbonyl]methyl]urea, Tris(trifluoroacetate) (19a). This compound was obtained as an oil (4.75 g, 86% yield) according to the procedure described for 13a and starting from 18a (6 g, 7.3 mmol). ¹H NMR (DMSO- d_6) δ : 1.2–1.65 (m, 12H), 1.9 (m, 2H), 2.9–3.15 (m, 12H), 3.6 (d, 2H), 6.1 (t, 1H), 6.3 (t, 1H), 6.8–9 (m, 11H). ¹³C NMR (DMSO- d_6) δ : 22.8, 23.7, 25.6, 25.8, 27.0, 28.3, 28.9, 36.1, 38.3, 38.5, 40.5, 42.7, 43.7, 46.5, 156.7, 157.9, 169.6.

Preparation of Compound 21a. 3-[(1,1-Dimethylethoxy)carbonylamino]-20-[(1,1-dimethylethoxy)carbonyl]-12,-15-dioxo-2,4,11,14,20,24-hexaaza pentacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (20a). A solution of isobutyl chloroformate (0.97 g, 7.1 mmol) in THF (5 mL) was added dropwise to a solution cooled to -30 °C of **11a** (2.7 g, 7.1 mmol) and N-methylmorpholine (0.72 g, 7.1 mmol) in THF (40 mL). The reaction medium was stirred for 0.5 h and a solution of 17a (2.9 g, 7.1 mmol) in THF (30 mL) was added. Stirring was maintained for 2 h at -30 °C and then for 24 h at room temperature. After filtration of the reaction mixture and evaporation of the filtrate under vacuum, the residue obtained was purified by flash chromatography on silica gel (EtOAc) to afford **20a** as an oil (1.9 g, 34% yield). ¹H NMR (CDCl₃) δ : 1.1-1.6 (m, 50H), 2.1-2.2 (t, 2H), 2.8-2.9 (q, 2H), 3.0-3.1 (m, 6H), 3.2-3.3 (q, 2H), 3.6 (d, 2H), 6.7-6.9 (bs, 1 H), 7.7 (t, 1H), 8.0 (t, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

5-[(3-Aminopropyl)amino]-*N*-**[2-[6-[(aminoiminomethyl)amino]hexylamino]-2-oxoethyl]pentanamide (21a).** This compound was obtained as an oil (0.7 g, 90%) according to the procedure described for **13a** and starting from **20a** (0.84 g, 1.1 mmol). ¹H NMR (DMSO- d_6) δ : 1.2–1.7 (m, 12H), 1.8–1.9 (m, 2H), 2.2 (t, 2H), 2.8–3.2 (m, 10H), 3.6 (d, 2H), 7.7 (t, 1H), 7.8–8.0 (m, 4H), 8.1 (t, 1H), 8.5–8.7 (m, 3H). ¹³C NMR (D₂O) δ : 22.8, 24.6, 25.8, 26.2, 26.3, 28.6, 29.0, 35.4, 37.4, 40.0, 41.9, 43.4, 45.2, 48.2, 157.5, 172.0, 177.4.

Preparation of Compound 23. 3-[[(1,1-Dimethylethoxy)carbonyl]amino]-21-[(1,1-dimethylethoxy)carbonyl]-1,2,-15-dioxo-16-oxa-2,4,11,14,21,25-hexaazahexacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (22). To a stirred solution of 8a (1.0 g, 2.89 mmol) and triethylamine (0.45 g, 4.5 mmol) in THF (20 mL) was added a solution of 4-nitrophenyl chloroformate (0.58 g, 2.89 mmol) in THF (5 mL). The reaction mixture was stirred for 15 h at room temperature, and a solution of 17a (1.2 g, 2.89 mmol) in THF (10 mL) was then added. The reaction mixture was heated to 40 °C and stirred for 5 h. After concentration of the reaction mixture under reduced pressure, the residue was purified by MPLC on silica gel (methylcyclohexane/EtOAc 7/3 and then EtOAc) to afford 22 (1.0 g, 44% yield) as a transparent oil. ¹H NMR (CDCl₃) δ : 1.3–1.9 (m, 50H), 3.05–3.55 (m, 10H), 3.85 (d, 2H), 4.10 (t, 2H), 4.7-5.3 (br s, 1H), 5.4-5.6 (br s, 1H), 6.0-6.2 (br s, 1H), 8.3-8.5 (br s, 1H), 11.5 (s, 1H).

[4-[(3-Aminopropyl)amino]butoxycarbonylamino]-*N*-[6-[(aminoiminomethyl)amino]hexyl]acetamide, Tris-(trifluoroacetate) (23). This compound was obtained in the form of a translucent white amorphous solid (0.78 g, 84% yield) according to the procedure described for 13a and starting from 22 (1.0 g, 1.25 mmol). ¹H NMR (DMSO- d_6) δ : 1.2–1.55 (m, 8H), 1.6–1.75 (m, 4H), 1.8–1.95 (m, 2H), 2.8–3.1 (m, 10H), 3.55 (d, 2H), 3.95 (t, 2H), 6.7–7.4 (m, 4H), 7.5 (t, 1H), 7.75– 8.0 (m, 4H), 8.5–8.65 (m, 3H). ¹³C NMR (D₂O) δ : 23.0, 24.5, 26.1, 26.2, 26.3, 28.5, 28.9, 31.0, 37.3, 39.9, 41.8, 45.2, 48.1, 65.7, 158.0, 159.0, 178.5.

Procedure for the Preparation of Compound 31. 31 was synthesized from 10-[(phenylmethoxy)carbonyl amino]-decanoic acid and **9** in four steps as follows:

18-[(1,1-Dimethylethoxy)carbonyl]-12-oxo-2,13,18,22tetraazatricosanedioic Acid, 23-(1,1-Dimethylethyl) Ester 1-Phenylmethyl Ester (28). To a stirred solution of 10-[(phenylmethoxy)carbonylamino]decanoic acid (0.92 g, 2.92 mmol) in CHCl₃ (30 mL) were added at 0 °C DCC (1.05 g, 5.1 mmol) and HOBT (69 mg, 0.51 mmol). The mixture was allowed to stand for half an hour at 0 °C. Then **9** (1.2 g, 3.47 mmol) in solution in CHCl₃ (20 mL) was added dropwise at 0 °C. After the addition was complete, the mixture was allowed to warm gradually to room temperature overnight. The solvent was evaporated off under reduced pressure and the oily residue was purified by flash chromatography on silica gel (hexane/ EtOAc 1/1 then EtOAc) as a viscous oil. ¹H NMR (CDCl₃) δ : 1.2–1.7 (m, 38H), 2.1 (t, 2H), 3.0–3.3 (m, 10H), 4.7–4.9 (br s, 1H), 5.1 (s, 2H), 5.2 (br s, 1H), 5.7 (br s, 1H), 7.3–7.4 (m, 5H).

10-Amino-*N***-[4-](1,1-dimethylethoxycarbonyl)-[3-](1,1dimethylethoxycarbonyl)amino]propyl]amino]butyl]decanamide (29).** A mixture of **28** (1.1 g, 1.69 mmol) and 10% palladium on carbon (0.1 g) in ethanol (20 mL) was stirred at room temperature and under a hydrogen atmosphere for 4 h at atmospheric pressure. The catalyst was then filtered off, and the organic phase was evaporated to give an oily residue which was purified by flash chromatography on silica gel (EtOAc then EtOAc/MeOH/NH₄OH/6/4/0.1) to afford **29** (0.78 g, 90% yield). ¹H NMR (CDCl₃) *3*: 1.2–1.7 (m, 38H), 2.1 (t, 2H), 2.3–2.5 (br s, 2H), 2.6–2.8 (br s, 2H), 3.0–3.3 (m, 8H), 5.1 (br s, 1H), 5.8 (br s, 1H).

3-[(1,1-Dimethylethoxy)carbonylamino]-20-[(1,1-dimethylethoxy)carbonyl]-14-oxo-2,4,15,20,24-pentaazapentacos-2-endioic Acid, Bis(1,1-dimethylethyl) Ester (30). 29 (0.78 g, 1.52 mmol) was added at room temperature to a stirred solution of *N*,*N*-bis(*tert*-butoxycarbonyl)-*S*-meth-ylisothiourea 1 (1.76 g, 6.0 mmol) in THF (20 mL). The reaction mixture was stirred at room temperature for 3 days. After evaporation of the solvent, the obtained residue was purified by flash chromatography on silica gel (hexane/EtOAc 1/1 then EtOAc) to afford **30** (0.67 g, 59% yield) as a colorless oil. ¹H NMR (CDCl₃) δ : 1.2–1.7 (m, 56H), 2.1 (t, 2H), 2.3–3.3 (m, 8H), 3.4 (q, 2H), 5.1 (br s, 1H), 5.7 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

10-(Aminoiminomethylamino)-*N*-[**4-**[(**3-aminopropy**])**amino]butyl]decanamide, Tris(trifluoroacetate) (31).** This compound was obtained as a white amorphous solid (0.46 g, 75.4% yield) according to the procedure described for **13a** and starting from **30** (0.67 g, 0.88 mmol). ¹H NMR (DMSO- d_6) δ : 1.2–1.35 (m, 10H), 1.4–1.6 (m, 8H), 1.8–1.9 (m, 2H), 2.0 (t, 2H), 2.8–3.1 (m, 10H),7.6 (t, 1H), 7.8–8.0 (m, 4H), 8.5–8.7 (br s, 3H). ¹³C NMR (D₂O) δ : 23.8, 24.5, 26.2, 26.4, 26.5, 28.6, 28.9, 29.0, 29.1, 29.2, 36.6, 37.3, 39.2, 42.0, 45.2, 48.2.

Procedure for the Preparation of Compound 35. 35 was synthesized from **9** and 3-[(1,1-dimethylethoxy)carbonyl-amino]-13-oxa-12-oxo-2,4,11-triazapentadec-2-enedioic acid, 1-(1,1-dimethylethyl) ester, **33** which was obtained in two steps as follows.

3-[(1,1-Dimethylethoxy)carbonylamino]-13-oxa-12-oxo-2,4,11-triazapentadec-2-endioic Acid, 1-(1,1-Dimethylethyl) Ester 15-Methyl Ester (32). A solution of 2 (18.78 g, 47.6 mmol) in toluene (700 mL) was added to a stirred solution of methyl [(phenoxycarbonyl)oxy] acetate (10.0 g, 47.6 mmol) in toluene (100 mL). The stirred reaction mixture was heated at 40 °C for 5 h. The solvent was evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (hexane/EtOAc 8/2) to afford 32 (11.8 g, 52% yield). ¹H NMR (CDCl₃) δ : 1.3–1.6 (m, 26H), 3.2 (q, 2H), 3.4 (q, 2H), 3.8 (s, 3H), 4.6 (s, 2H), 4.8–5.0 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

3-[(1,1-Dimethylethoxy)carbonylamino]-13-oxa-12-oxo-2,4,11-triazapentadec-2-enedioic Acid, 1-(1,1-Dimethylethyl) Ester (33). This compound was obtained (8.9 g, 78.5% yield) as an oil by following a procedure analogous to preparation of **4a** and starting from **32** (11.7 g, 24.7 mmol). ¹H NMR (CDCl₃) δ : 1.3–1.7 (m, 26H), 3.2–3.4 (m, 4H), 4.5 (s, 2H), 4.9 (t, 1H), 8.4 (s, 1H), 11.0–11.5 (br s, 1H).

3-[(1,1-Dimethylethoxy)carbonylamino]-21-[(1,1-dimethylethoxy)carbonyl]-13-oxa-12,15-dioxo-2,4,11,16,21,-25-hexaazacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (34). To a stirred solution of 33 (3.0 g, 6.52 mmol) in CH₂Cl₂ (60 mL) were added at 0 °C DCC (2.7 g, 13.1 mmol) and HOBT (0.176 g, 1.31 mmol). This mixture was allowed to stand for 0.5 h at 0 °C. Then 9 (2.25 g, 6.52 mmol) in solution in CH₂Cl₂ (30 mL) was added dropwise at 0 °C. After the addition was complete, the mixture was allowed to warm gradually to room temperature and left to stand 2 days. The solvent was evaporated off under reduced pressure and the obtained residue was purified by flash chromatography on silica gel (methylcyclohexane/EtOAc 3/7) to afford 34 (4.5 g, 87.8% yield) as a viscous oil. ¹H NMR (CDCl₃) δ : 1.3–1.8 (m, 50H), 3.1-3.5 (m, 12H), 4.5 (s, 2H), 4.8-5.0 (br s, 1H), 5.2-5.4 (br s, 1H), 6.2-6.6 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

[6-(Aminoiminomethylamino)hexyl]carbamic Acid, 2-[[4-[(3-Aminopropyl)amino]-butyl]amino]-2-oxoethyl Ester, Tris(trifluoroacetate) (35). This compound was obtained in the form of a translucent white amorphous solid (4.0 g, 84.7% yield) according to the procedure described for **13a** and starting from **34** (4.5 g, 5.72 mmol). ¹H NMR (DMSO-*d*₆) δ : 1.2–1.6 (m, 12H), 1.8–2.0 (m, 2H), 2.3–3.2 (m, 12H), 4.3 (s, 2H), 7.2 (t, 1H), 7.6 (t, 1H), 7.8–8.0 (m, 4H), 8.5–8.7 (br s, 3H). ¹³C NMR (D₂O) δ : 23.6, 24.5, 26.2, 26.3, 26.4, 28.5, 29.4, 37.3, 38.9, 41.8, 45.2, 48.1, 63.7, 157.9, 158.2, 172.0.

Procedure for the Preparation of Compound 39. 39 was synthesized from **9** and carbobenzyloxyglycine in four steps as follows.

10-[(1,1-Dimethylethoxy)carbonyl]-4-oxo-2,5,10,14-tetraazapentadecanedioic Acid, 1-Phenylmethyl 15-(1,1-Dimethylethyl) Ester (36). To a stirred solution of carbobenzyloxyglycine (7.06 g, 34.0 mmol) in CHCl₃ (100 mL) were added DCC (7.0 g, 34 mmol) and HOBT (0.95 g, 7 mmol). The mixture was allowed to stand for half an hour at 0 °C. Then 9 (9.32 g, 27 mmol) in solution in CHCl₃ (100 mL) was added dropwise at 0 °C. After the addition was complete, the mixture was allowed to warm gradually to room temperature overnight. The solvent was evaporated off under reduced pressure, and the oily residue was purified by flash chromatography on silica gel (methylcyclohexane/EtOAc 1/1 then EtOAc) to afford 36 (12.5 g, 86% yield) as a white solid. ¹H NMR (CDCl₃) δ : 1.4– 1.8 (m, 24H), 3.0-3.4 (m, 8H), 3.8 (d, 2H), 4.8-5.0 (br s, 1H), 5.1 (s, 2H),5.3-5.7 (br s, 1H), 6.1-6.6 (br s, 1H), 7.3-7.4 (m, 5H).

[3-[[(1,1-Dimethylethoxy)carbonyl][4-[2-amino-1-oxoethyl)amino]butyl]amino]propyl]carbamic Acid, (1,1-Dimethylethyl) Ester (37). A mixture of 36 (12.5 g, 23.32 mmol) and 10% palladium on carbon (0.88 g) in ethanol (200 mL) was stirred at room temperature under a hydrogen atmosphere for 5 h at atmospheric pressure. The catalyst was then filtered off, and the organic phase was evaporated to give 37 (9.3 g, quantitative yield) in the form of an oily residue which was used without further purification for the preparation of 38. ¹H NMR (CDCl₃) δ : 1.4–1.8 (m, 24H), 1.9–2.0 (br s, 2H), 3.0–3.4 (m, 10H), 4.5 (br s, 1H), 5.3–5.4 (br s, 1H).

3-[(1,1-Dimethylethoxy)carbonylamino]-21-[(1,1-dimethylethoxy)carbonyl]-12,15-dioxo-2,4,11,13,16,21,25heptaazahexacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (38). 4-Nitrophenyl chloroformate (0.5 g, 2.5 mmol) was added in small portions to a solution of 37 (1.0 g, 2.5 mmol) and triethylamine (2.5 mmol) in anhydrous THF (50 mL). The reaction medium was stirred for 1 h at room temperature, and then a solution of 2 (0.9 g, 2.5 mmol) in anhydrous THF (20 mL) was added dropwise. Stirring was continued overnight at room temperature, and the solvent was evaporated off under reduced pressure. The obtained residue was purified by flash chromatography on silica gel (EtOAc, MeOH 9.5/0.5) to afford 38 (1.1 g, 56% yield). ¹H NMR (CDCl₃) δ : 1.2–1.7 (m, 50H), 2.8–3.1 (m, 12H), 3.2 (q, 2H), 5.9 (t, 1H), 6.1 (t, 1H), 6.7–6.8 (br s, 1H), 7.8 (t, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

N-[6-(Aminoiminomethylamino)hexyl]-*N*-[2-[[4-[(3-aminopropyl)amino]butyl]amino]-2-oxoethyl]urea, Tris(trifluoroacetate) (39). This compound was obtained in the form

of an oil (0.58 g, 53% yield) according to the procedure described for **13a** and starting from **38** (1.2 g, 1.5 mmol). ¹H NMR (D₂O) δ : 1.2–1.7 (m, 12H), 1.9–2.1 (m, 2H), 3.0–3.2 (m, 14H). ¹³C NMR (D₂O) δ : 23.8, 24.6, 26.3, 26.4, 26.5, 28.7, 29.7, 37.6, 38.3, 40.3, 41.9, 44.1, 45.1, 48.1, 157.8, 161.5, 174.3.

Procedure for the Preparation of Compound 42. 42 was synthesized from **2** and thioglycolic acid in three steps as follows.

[6[(2-Mercapto-1-oxoethyl)amino]hexyl]carbonimidoyl]biscarbamic Acid, Bis(1,1-dimethylethyl) Ester (40). Carbonyldiimidazole (1.4 g, 8.4 mmol) was added in small portions to a solution of thioglycolic acid (0.55 mL, 7.7 mmol) in CH₂-Cl₂ (30 mL). The reaction medium was stirred overnight at room temperature, and then a solution of **2** (2.5 g, 7 mmol) in CH₂Cl₂ (20 mL) was added dropwise. Stirring was continued for 5 h at room temperature, and the solvent was evaporated off under reduced pressure. The obtained residue was purified by flash chromatography on silica gel (EtOAc/cyclohexane 1/1) to afford **40** as an oil (1.5 g, 55% yield). ¹H NMR (CDCl₃) δ : 1.3–1.7 (m, 26H), 1.9 (t, 1H), 3.2–3.4 (m, 4H), 3.5–3.6 (q, 2H), 6.6–6.8 (br s, 1H),8.3 (t, 1H), 11.5 (s, 1H).

3-[(1,1-Dimethylethoxy)carbonylamino]-21-[(1,1-dimethylethoxy)carbonyl]-12,15-dioxo-14-thia-2,4,11,16,-21,25-hexaazahexacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (41). To a stirred solution of 40 (1.37 g, 3.5 mmol) and triethylamine (0.7 g, 7.0 mmol) in THF (25 mL) was added a solution of 4-nitrophenyl chloroformate (0.7 g, 3.5 mmol) in THF (5 mL). The reaction mixture was stirred for 1 h at room temperature, and a solution of 9 (1.2 g, 3.5 mmol) in THF (10 mL) was then added. Stirring was continued for one night at room temperature. After concentration of the reaction medium under reduced pressure, the residue was purified by flash chromatography on silica gel (EtOAc/cyclohexane 6/4) to afford 41 (2.0 g, 71% yield) as a transparent oil. ¹H NMR (CDCl₃) δ: 1.3–1.8 (m, 50H), 3.1–3.4 (m, 12H), 3.5 (s, 2H), 4.7-5.3 (br s, 1H), 6.7-6.8 (br s, 1H), 7.0-7.2 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

[4-[(3-Aminopropyl)amino]butyl]carbamothioic Acid, *S*-[2-[[6-(Aminoiminomethylamino)hexyl]amino]-2-oxoethyl Ester, Tris(trifluoroacetate) (42). This compound was obtained in the form of a translucent white amorphous solid (1.45 g, 77% yield) according to the procedure described for 13a and starting from 41 (2 g, 2.5 mmol). ¹H NMR (DMSO d_6) δ : 1.2–1.7 (m, 12H), 1.8–2.0 (m, 2H), 2.9–3.2 (m, 12H), 3.5 (s, 2H), 7.5 (t, 1H), 7.8–8.0 (m, 5H), 8.3 (t, 1H), 8.5–8.6 (br s, 2H). ¹³C NMR (D₂O) δ : 23.7, 24.5, 26.1, 26.3, 26.5, 28.5, 28.8, 33.8, 37.3, 40.1, 41.1, 41.8, 45.2, 48.1, 157.5, 169.1, 171.8.

Procedure for the Preparation of Compound 46. 46 was synthesized from **9** and methyl 2-(chlorosulfonyl)acetate in four steps as follows.

9-[(1,1-Dimethylethoxy)carbonyl]-3-thia-3,3-dioxo-4,9,-13-triazatetradecanedioic Acid, 14-(1,1-Dimethylethyl) 1-Methyl Ester (43). To a stirred solution of **9** (6.2 g, 18 mmol) and triethylamine (2.8 mL, 20 mmol) in CH₂Cl₂ (40 mL) cooled to -20 °C was added dropwise a solution of methyl 2-(chlorosulfonyl)acetate in CH₂Cl₂ (20 mL). The stirring was continued at -20 °C for 10 min, and the mixture was then allowed to warm to room temperature for 4 h. The solvent was evaporated off under reduced pressure, and the oily residue was purified by flash chromatography on silica gel (cyclohexane/EtOAc 1/1) to afford **43** (3.54 g, 41% yield). ¹H NMR (CDCl₃) δ : 1.4–1.7 (m, 24H), 3.0–3.3 (m, 8H), 3.8 (s, 3H), 4.0 (s, 2H), 4.6–5.3 (br s, 2H).

9-[(1,1-Dimethylethoxy)carbonyl]-3-thia-3,3-dioxo-4,9,-13-triazatetradecanedioic Acid, 14-(1,1-Dimethylethyl) Ester (44). This compound was obtained (2.9 g, 85% yield) as a yellow oil by following a procedure analogous to preparation of **4a** and starting from **43** (3.5 g, 7.3 mmol). ¹H NMR (CDCl₃) δ : 1.4–1.8 (m, 24H), 3.0–3.3 (m, 8H), 4.0 (s, 2H), 4.7–5.4 (br s, 2H).

3-[(1,1-Dimethylethoxy)carbonylamino]-20-[(1,1-dimethylethoxy)carbonyl]-14-thia-14,14-dioxo-2,4,11,15,-20,24-hexaazapentacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (45). To a stirred solution of 44 (2.7 g, 5.8 mmol) in CH₂Cl₂ (30 mL) was added carbonyldiimidazole (1.2 g, 6.5 mmol) in small portions and at room temperature. The reaction mixture was stirred for 3 h. Then **2** (2.28 g, 5.8 mmol) and triethylamine (0.9 mL, 6.0 mmol) in solution in CH₂Cl₂ (20 mL) were added dropwise at room temperature. After the addition was complete, stirring was continued overnight. The solvent was evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (EtOAc/cyclohexane 1/1 then 4/1) to give **45** (2.17 g, 46% yield). ¹H NMR (CDCl₃) δ : 1.3–1.7 (m, 50H), 3.0–3.5 (m, 12H), 3.9 (s, 2H), 4.7–5.0 (br s, 1H), 5.1–5.3 (br s, 1H), 6.4–6.6 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

2-[[4-[(3-Aminopropyl)amino]butyl]aminosulfonyl]-*N*-**[6-aminoiminomethylamino)hexyl]-acetamide, Tris(tri-fluoroacetate) (46).** This compound was obtained as an oil (1.47 g, 73% yield) according to the procedure described for **13a** and starting from **45** (2.17 g, 2.7 mmol). ¹H NMR (DMSO- d_6) δ : 1.2–1.7 (m, 12H), 1.9–2.0 (m, 2H), 2.9–3.2 (m, 12H), 3.9 (s, 2H), 7.2 (t, 1H), 7.7 (t, 1H), 7.8–8.0 (m, 4H), 8.2 (t, 1H), 8.6–8.7 (br s, 2H). ¹³C NMR (D₂O) δ : 23.2, 24.3, 26.0, 26.1, 27.0, 28.3, 28.5, 37.2, 40.8, 41.7, 42.9, 45.0, 47.8, 58.1, 157.4, 173.3.

Procedure for the Preparation of Compound 50. 50 was synthesized from **6** and 1-(1,1-dimethylethyl)-6-[(1,1-dimethylethoxy)carbonyl]-12-oxo-2,6,11,triazatetradecane dioate **48** obtained in two steps as follows.

6-[(1,1-Dimethylethoxy)carbonyl]-12-oxo-2,6,11-triazatetradecanedioic Acid, 1-(1,1-Dimethylethyl)-14-ethyl Ester (47). To a stirred solution of ethyl hydrogen malonate (1.60 g, 12.0 mmol) in CHCl₃ (50 mL) were added at 0 °C DCC (2.50 g, 12.0 mmol) and HOBT (0.13 g, 1 mmol). The mixture was allowed to stand for half an hour at 0 °C. Then **5** (3.12 g, 9.0 mmol) in solution in CHCl₃ (20 mL) was added dropwise at 0 °C. After the addition was complete, the mixture was allowed to warm gradually to room temperature overnight. The solvent was evaporated off under reduced pressure, and the oily residue was purified by flash chromatography on silica gel (EtOAc/hexane 1/1 then EtOAc) to afford **47** (2.67 g, 69%) as a viscous oil. ¹H NMR (CDCl₃) δ : 1.25 (t, 3H), 1.4–1.7 (m, 24H), 3.10–3.35 (m, 10H), 4.2 (q, 2H).

6-[(1,1-Dimethylethoxy)carbonyl]-12-oxo-2,6,11-triazadecanedioic Acid, 1-(1,1-Dimethylethyl) Ester (48). This compound was obtained (0.75 g, 84% yield) as an oil by following a procedure analogous to preparation of 4a and starting from 47 (0.95 g, 2.0 mmol). ¹H NMR (DMSO- d_6) δ : 1.40 (s, 18H), 1.55 (m, 6H), 2.9 (m, 4H), 3.15 (m, 6H).

3-[(1,1-Dimethylethoxycarbonyl)amino]-11-methyl-20-[1,1-dimethylethoxycarbonyl)-12,14-dioxo-2,4,11,15,20,-24-hexaazapentacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (49). To a stirred solution of 48 (1.2 g, 2.7 mmol) in CH₂Cl₂ (50 mL) were added at 0 °C DCC (1.2 g, 5.4 mmol) and HOBT (0.2 g, 1.5 mmol). The mixture was allowed to stand 0.5 h at 0 °C. Then 6 (1.0 g, 2.7 mmol) in solution in CH₂Cl₂ (30 mL) was added dropwise at 0 °C. After the addition was complete, the mixture was allowed to warm gradually to room temperature and left to stand one night. The solvent was evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (EtOAc/EtOH 95/5) to afford 49 (1.8 g, 83% yield) as a viscous oil. ¹H NMR (CDCl₃) δ : 1.3–1.7 (m, 50H), 2.9–3.3 (m, 15H), 3.4 (q, 2H), 4.8 and 5.3 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

N-Methyl-N-[6-(aminoiminomethylamino)hexyl]-N-[4-[(3 aminopropyl)amino]butyl]-malonodiamide (50). This compound was obtained in the form of a translucent white amorphous solid (1.44 g, 85% yield) according to the procedure described for **13a** and starting from **49** (1.8 g, 2.3 mmol). ¹H NMR (DMSO-*d*₆) δ : 1.2–1.35 (m, 4H), 1.4–1.7 (m, 8H), 1.5– 2.0 (m, 2H), 2.8–3.0 (m, 9H), 3.05–3.15 (m, 4H), 3.2–3.3 (m, 4H), 7.5–7.65 (br s, 1H), 7.75–7.9 (br s, 3H), 8.0–8.1 (br s, 1H), 8.4–8.6 (br s, 3H). ¹³C NMR (D₂O) δ : 23.7, 24.5, 26.3, 26.4, 26.9, 28.0, 28.5, 36.6, 37.3, 39.5, 41.8, 45.2, 48.1, 48.7, 51.5, 157.5, 170.0, 170.5.

Procedure for the Preparation of Compound 55. 55 was synthesized from **2** and 3-[[4-[[(1,1-dimethylethoxy)car-

bonyl][3-[[(1,1-dimethylethoxy)carbonyl]amino]propyl]amino]butyl] [(1,1-dimethylethoxy)carbonyl]amino]propanoic acid **53** which was obtained in three steps as follows.

3-[[4-[[(1,1-Dimethylethoxy)carbonyl][3-[[(1,1-dimethylethoxy)carbonyl]amino]propyl]amino]butyl]amino] propanoic Acid, Methyl Ester (51). Methylacrylate (0.5 g, 5.8 mmol) in solution in methanol (30 mL) was added with stirring to a solution of 9 (2.0 g, 5.8 mmol) in methanol (20 mL). The reaction was refluxed for 3 h. The solvent was evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (EtOAc then EtOAc/EtOH 6/4) to afford 51 (0.98 g, 39%). ¹H NMR (CDCl₃) δ : 1.4–1.9 (m, 25H), 2.9–3.3 (m, 12H), 3.72 (s, 3H), 4.7–5.4 (br s, 1H).

3-[[4-[[(1,1-Dimethylethoxy)carbonyl][3-[[(1,1-dimethylethoxy)carbonyl]amino]propyl]amino]butyl][(1,1-dimethylethoxy)carbonyl]amino]propanoic Acid, Methyl Ester (52). To a stirred solution of 51 (0.86 g, 2 mmol) and triethylamine (0.14 g, 1.4 mmol) in THF (30 mL) was added a solution of diterbutyl dicarbonate (0.48 g, 2 mmol) in THF (20 mL). The reaction mixture was stirred for 2 h at room temperature. Then the solvent was evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (EtOAc/cyclohexane 5/5) to afford 52 (0.9 g, 85%). ¹H NMR (CDCl₃) δ : 1.4–1.8 (m, 33H), 2.58 (t, 2H), 3.3–3.3 (m, 8H), 3.45 (t, 2H), 3.68 (s, 3H), 4.7–5.4 (br s, 1H).

3-[[4-[[(1,1-Dimethylethoxy)carbonyl][3-[[(1,1-dimethylethoxy)carbonyl]amino]propyl]amino]butyl][(1,1-dimethylethoxy)carbonyl]amino]propanoic Acid (53). This compound was obtained (0.74 g, 90% yield) as a yellow oil by following a procedure analogous to preparation of **4a** and starting from **52** (0.34 g, 1.6 mmol). ¹H NMR (CDCl₃) δ : 1.4–1.8 (m, 33H), 2.61 (t, 2H), 3.0–3.3 (m, 8H), 3.5 (t, 2H), 4.7–5.4 (br s, 1H).

3-[(1,1-Dimethylethoxy)carbonylamino]-15,20-bis[(1,1dimethylethoxy)carbonyl]-12-oxo-2,4,11,15,20,24-hexaazapentacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (54). A solution of isobutylchloroformate (0.165 mL, 1.3 mmol) in THF (5 mL) was added dropwise to a solution, cooled to 30 °C, of 53 (0.66 g, 1.3 mmol) and N-methylmorpholine (0.28 mL, 2.6 mmol) in THF (30 mL). The reaction medium was stirred for 0.5 h, and a solution of 2 (0.46 g, 1.3 mmol) in THF (20 mL) was added dropwise. Stirring was maintained for 2 h at -30 °C and then overnight at room temperature. After filtration of the reaction medium and evaporation of the filtrate in vacuo, the residue obtained was purified by flash chromatography on silica gel (EtOAc/cyclohexane 8/2) to afford 54 (0.9 g, 66% yield). ¹H NMR (CDCl₃) δ : 1.3–1.8 (m, 59H), 2.43 (t, 2H), 3.0-3.3 (m, 10H), 3.4 (t, 2H), 3.46 (t, 2H), 4.7-5.4 (br s, 1H), 6.2-6.4 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

3-[[4-[(3-Aminopropyl)amino]butyl]amino]-*N***-[6-(aminoiminomethylamino)hexyl]-propanamide, Tetrakis(tri-fluoroacetate) (55).** This compound was obtained in the form of an oil (0.6 g, 74% yield) according to the procedure described for **7a** and starting from **54** (0.85 g, 1 mmol). ¹H NMR (DMSO-*d*₆) δ: 1.2–1.5 (m, 8H), 1.55–1.7 (m, 4H), 1.8–2.0 (m, 2H), 2.5 (t, 2H), 2.8–3.2 (m, 14H), 7.67 (t, 1H), 7.8–8.0 (br s, 3H), 8.12 (t, 1H), 8.5–8.65 (br s, 2H), 8.7–8.85 (br s, 2H). ¹³C NMR (D₂O) δ: 23.4, 23.5, 24.5, 26.2, 26.4, 28.5, 28.8, 31.8, 37.3, 40.2, 41.9, 44.3, 45.3, 47.6, 47.8, 157.4, 170.3.

Procedure for the Preparation of Compound 60. This compound was synthesized starting from [(6-oxohexyl)carboimidoyl]biscarbamic acid, bis(1,1-dimethylethyl) ester, **56** and *N*-[4[(2 aminoethoxy)carbonylamino]butyl]-1,3-propanebiscarbamic acid, bis(1,1-dimethyl ethyl ester, **58** obtained, respectively, in one step from [(6-aminomethylhexyl)carboimidoyl]biscarbamic acid, bis(1,1-dimethylethyl) ester, **6** and in two steps from carbamic acid, (2-hydroxyethyl)-phenylmethyl ester as follows.

[(6-Oxohexyl)carboimidoyl]biscarbamic Acid, Bis(1,1-dimethylethyl) Ester (56). Pyridinium dichromate (2.6 g, 7.0 mmol) was added in small portions to a solution of **5** (1.4 g, 3.9 mmol) in CH₂Cl₂ (50 mL). The reaction medium was stirred

for 1 day at room temperature, then filtered, and washed with CH₂Cl₂ (3 \times 25 mL). The filtrates were evaporated off under reduced pressure, and the obtained residue was purified by flash chromatography on silica gel (EtOAc/methylcyclohexane 2.5/7.5) to afford **56** (0.42 g, 30%) as a colorless oil. ¹H NMR (CDCl₃) δ : 1.3–1.8 (m, 24H), 2.43 (t, 2H), 3.42 (t, 2H), 8.3 (t, 1H), 9.77 (t, 1H), 11.5 (s, 1H).

12-[1,1-Dimethylethoxycarbonyl]-5-oxa-6-oxo-2,7,12,-16-tetraazaheptadecanedioic Acid, 1-(Phenylmethyl)-17-(1,1-Dimethylethyl) Ester (57). To a stirred solution of carbamic acid (2-hydroxyethyl)-phenylmethyl ester (2.0 g, 10.25 mmol) and triethylamine (2.0 g, 20.5 mmol) in anhydrous toluene (100 mL) was added dropwise at room temperature a solution of phenylchloroformate (2.8 g, 10.25 mmol) in toluene (20 mL). The reaction mixture was stirred for 2 h at room temperature, and a solution of 9 (3.53 g, 10.25 mmol) in toluene (100 mL) was then added. The reaction mixture was heated to 60 °C and stirred for 16 h. After evaporation of the solvent, the obtained residue was purified by flash chromatography on silica gel (methylcyclohexane/EtOAc 8/2) to afford 57 (3.3 g, 57%). ¹H NMR (CDCl₃) δ : 1.4–1.8 (m, 24H), 3.0–3.3 (m, 8H), 3.4-3.5 (m, 2H), 4.1-4.2 (m, 2H), 4.7-5.0 (br s, 2H), 5.10 (s, 2H), 5.15-5.4 (br s, 1H), 7.3-7.4 (m, 5H).

N-[4[(2-Aminoethoxy)carbonylamino]butyl]-1,3-propane-biscarbamic Acid, Bis(1,1-dimethylethyl) Ester (58). A mixture of 57 (3.3 g, 5.8 mmol) and 10% palladium on carbon (0.4 g) in ethanol (70 mL) was stirred at room temperature under a hydrogen atmosphere for 5 h at atmospheric pressure. The catalyst was then filtered off and the organic phase was evaporated to give 58 (2.45 g, 97% yield) in the form of an oily residue which was used without further purification for the preparation of 59. ¹H NMR (CDCl₃) δ : 1.4–1.8 (m, 26H), 2.9 (t, 2H), 3.0–3.4 (m, 8H), 4.08 (t, 2H), 4.6–5.4 (br s, 2H).

3-[(1,1-Dimethylethoxycarbonylamino]-21-[1,1-dimethylethoxycarbonyl)-14-oxa-15-oxo-2,4,11,16,21,25-hexaazahexacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (59). To a stirred solution of 56 (0.43 g, 1.2 mmol) and 59 (0.52 g, 1.2 mmol) in EtOH (40 mL) acidified to pH 6 with AcOH was added in small portions NaBH₃CN (0.15 g, 2.5 mmol). The reaction medium was stirred for 1 day. After concentration of the reaction medium under reduced pressure, the residue was taken up in $CH_2Cl_2-H_2O$ (100 mL), and the aqueous phase was extracted with CH₂Cl₂. The combined organic layers were dried, concentrated, and purified by flash chromatography on silica gel (EtOAc then EtOAc/EtOH/NH4OH 8/2/0.02) to afford **59** (0.45 g, 48% yield) as a yellow oil. ¹H NMR (CDCl₃) δ : 1.2– 2.2 (m, 50H), 2.7 (t, 2H), 2.8-3.0 (m, 2H), 3.05-3.35 (m, 8H), 3.5 (q, 2H), 4.2 (t, 2H), 4.8 (br s, 1H), 5.0 (br s, 1H), 5.3 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

[4-[(3-Aminopropyl)amino]butyl]carbamic Acid, 2[[6-(Aminoiminomethylamino)hexyl]amino]ethyl Ester (60). This compound was obtained in the form of a translucent white amorphous solid (0.4 g, 44% yield) according to the procedure described for **13a** and starting from **59** (0.85 g, 1.1 mmol). ¹H NMR (DMSO- d_6) δ : 1.2–1.4 (m, 4H), 1.45–1.7 (m, 8H), 1.8– 2.0 (m, 2H), 2.8–3.3 (m, 14H), 4.17 (t, 2H), 7.29 (t, 1H), 7.71 (t, 1H), 7.8–8.0 (br s, 3H), 8.5–8.8 (br s, 4H). ¹³C NMR (D₂O) δ : 23.7, 24.5, 26.0, 26.1, 26.7, 28.4, 37.3, 40.6, 41.8, 45.2, 47.3, 48.0, 48.2, 48.5, 61.1, 158.4, 170.5.

Procedure for the Preparation of Compound 68. This compound was synthesized with *N*,*N*-bis(*tert*-butoxycarbonyl)-*S*-methylisothiourea **1** and 22-amino-6[(1,1-dimethylethoxy)carbonyl]-12,16-dioxo-11-oxa-2,6,13,15-tetraaza-docosanoic acid, (1,1-dimethylethyl) ester **66** obtained in six steps from *N*-(6bromohexanoyl)glycine, ethyl ester and **8a**.

N-(6-Cyanohexanoyl)glycine, Ethyl Ester (61). Powdered potassium cyanide (6.5 g, 100 mmol) was added in small portions to a solution of *N*-(6-bromohexanoyl)glycine, ethyl ester (23.73 g, 84.7 mmol) in EtOH (200 mL). The reaction medium was refluxed for 15 h. After concentration of the mixture under reduced pressure, the residue was taken up with CH_2Cl_2 , and the organic phase was washed with a brine solution. The organic phase was dried and concentrated under reduced pressure to give **61** (19.0 g, 99%) which was used without purification. ¹H NMR (CDCl₃) δ : 1.29 (t, 3H), 1.45–1.55 (m, 2H), 1.6–1.8 (m, 4H), 1.27 (t, 2H), 2.36 (t, 2H), 4.03 (d, 2H), 4.22 (q, 2H), 5.35–6.05 (br s, 1H).

N-(6-Cyanohexanoyl)glycine (62). This compound was obtained (10.2 g, 61% yield) as a yellow oil by following a procedure analogous to preparation of **4a** and starting from **61** (19.0 g, 84.0 mmol). ¹H NMR (CDCl₃) δ : 1.45–1.55 (m, 2H), 1.6–1.8 (m, 4H), 2.3 (t, 2H), 2.36 (t, 2H), 4.08 (d, 2H), 6.15–6.25 (br s, 1H).

N-(7-Aminoheptanoyl)glycine, Sodium Salt (63). A mixture of *N*-(6-cyanohexanoyl)glycine (8.2 g, 41.4 mmol) and Raney nickel (800 mg) in ethanol (100 mL) and 1 M NaOH solution (60 mL) was stirred at room temperature under a hydrogen atmosphere and under a pressure of 3.5. 10^5 Pa for 8 h. Then 1 M HCl (20 mL) was added, and the mixture was concentrated under reduced pressure to give **63** (10.2 g) as a white pasty solid which contains the expected salt and sodium chloride and which can be used without further purification in the next step. ¹H NMR (DMSO-*d*₆) δ : 1.15–1.6 (m, 8H), 2.08 (t, 2H), 2.56 (t, 2H), 3.32 (d, 2H), 7.1–7.25 (br s, 1H).

N-[7-[(Phenylmethoxycarbonyl)amino]heptanoyl]glycine (64). To a stirred solution of 63 (2 g, 9.9 mmol) in EtOH (50 mL) and water (50 mL) were added sodium carbonate (1.0 g, 10 mmol) and then dropwise benzylchloroformate (2.3 g, 19.8 mmol). After stirring overnight at room temperature, the reaction medium was brought to pH 1 with 1 M hydrochloric acid and then extracted with CH_2Cl_2 (3 × 100 mL). The organic layers were combined, dried, and concentrated under reduced pressure, and the obtained crude product was purified by flash chromatography on silica gel (EtOAc then EtOAc/EtOH/NH₄-OH 6/3/0.5) to afford 64 (1.3 g, 40% yield) as a white pasty solid. ¹H NMR (DMSO- d_6) δ : 1.2–1.6 (m, 8H), 2.09 (t, 2H), 2.98 (td, 2H), 3.64 (d, 2H), 5.01 (s, 2H), 7.25 (t, 1H), 7.3–7.4 (m, 5H), 7.94 (t, 1H).

19-[(1,1-Dimethylethoxy)carbonyl]-9,13-dioxo-14-oxa-2,10,12,19,23-pentaza-tetracosanedioic Acid, 1-(Phenylmethyl) 24-(1,1-Dimethylethyl) Ester (65). To a stirred solution of $\mathbf{64}$ (0.5 g, 1.49 mmol) and triethylamine (0.18 g, 1.8 mmol) in THF (25 mL) was added dropwise at 0 °C a solution of diphenylphosphoryl azide (0.46 g, 1.67 mmol). The reaction mixture was stirred at room temperature for 45 min. After concentration of the mixture under reduced pressure, the residue was taken up with toluene (20 mL). To this solution were added 8a (1.04 g, 3.0 mmol) and triethylamine (0.18 g, 1.8 mmol). The solution medium was stirred at reflux for 24 h. After evaporation of the solvent, the obtained residue was purified by flash chromatography on silica gel (methylcyclohexane/EtOAc 1/1 to 1/9) to afford 65 (0.4 g, 40% yield). ¹H NMR (CDCl₃) δ: 1.2–1.7 (m, 32H), 2.15 (t, 2H), 3.05–3.3 (m, 8H), 4.05 (t, 2H), 4.5 (t, 2H), 4.7-4.9 (br s, 1H), 5.08 (s, 2H), 5.7-5.9 (br s, 1H), 6.4-6.6 (br s, 1H), 7.3-7.4 (m, 6H).

22-Amino-6-[(1,1-dimethylethoxy)carbonyl]-12,16-dioxo-11-oxa-2,6,13,15-tetraaza-docosanoic Acid, (1,1-Dimethylethyl) Ester (66). A mixture of **65** (0.33 g, 0.48 mmol) and 5% palladium on carbon (0.1 g) in ethanol (15 mL) was stirred at room temperature and under a hydrogen atmosphere for 5 h at atmospheric pressure. The catalyst was then filtered off, and the organic phase was evaporated to give **66** (0.26 g, quantitative yield) in the form of an oily residue which was used without further purification for the preparation of **67**. ¹H NMR (CDCl₃) &: 1.2–1.75 (m, 32H), 2.2 (t, 2H), 2.8 (t, 2H), 2.85–3.3 (m, 8H), 4.07 (t, 2H), 4.53 (t, 2H), 4.7–5.4 (br s, 1H), 5.7–6.1 (br s, 1H), 6.4–7.1 (br s, 1H).

3-[[(1,1-Dimethylethoxyl)carbonyl]amino]-21-[(1,1-dimethylethoxy)carbonyl]-11,15-dioxo-16-oxa-2,4,12,14,21,-25-hexaazahexacos-2-enedioic Acid, Bis(1,1-dimethylethyl) Ester (67). 66 (0.26 g, 0.48 mmol) was added at room temperature to a stirred solution of *N***,***N***-bis(***tert***-butoxycarbonyl)-***S***-methylisothiourea 1 (0.28 g, 0.95 mmol) in THF (20 mL). The reaction medium was stirred at room temperature for 4 days. After evaporation of the solvent, the obtained residue was purified by flash chromatography on silica gel (methylcyclohexane/EtOAc 4/6) to afford 67 (0.10 g, 27% yield)** as a colorless oil. ¹H NMR (CDCl₃) δ : 1.0–1.8 (m, 50H), 2.16 (t, 2H), 3.05–3.3 (m, 6H), 3.4 (q, 2H), 4.07 (t, 2H), 4.55 (t, 2H), 5.2–5.3 (br s, 1H), 5.7–5.8 (br s, 1H), 6.45–6.55 (br s, 1H), 8.3 (t, 1H), 11.5 (s, 1H).

7-[(Aminoiminomethyl)amino]-*N*-**[[4-[(3-aminopropyl)amino]butoxy]carbonylaminomethyl]heptanamide, Tris-(trifluoroacetate) (68).** This compound was obtained (0.050 g, 54% yield) in the form of a translucent white amorphous solid according to the procedure described for **13a** and starting from **67** (0.1 g, 0.127 mmol). ¹H NMR (DMSO-*d*₆) δ : 1.2–1.35 (m, 4H), 1.4–1.7 (m, 8H), 1.8–1.95 (m, 2H), 2.06 (t, 2H), 2.8– 3.1 (m, 8H), 3.96 (t, 2H), 4.32 (t, 2H), 6.85–7.4 (br s, 3H), 7.55– 7.7 (m, 2H), 7.8–8.0 (m, 4H), 8.33 (t, 1H), 8.55–8.7 (m, 2H).

Biology. Experimental graft-versus-host disease was induced in cyclophosphamide immunosuppressed B6D2F1 mice²⁴ by intravenous injection of 2×10^8 viable spleen cells from B6 origin. Generally, untreated control animals died between days 14 and 21 post-GVHD induction. Compounds were dissolved in aqueous solution and administered from days 1 to 10 (day 6 omitted) by intraperitoneal route. In such experimental conditions, control vehicles were unable to modify survival. Survival was followed until day 60 post-cell injection.

Heterotopic abdominal heart transplant was done according to the method of Ono and Lindsey.³² End-to-side anastomoses of the aorta and pulmonary artery of the donor heart (Dark Agouti) to the recipient (Lewis rat) abdominal aorta and vena cava were performed. Graft function was assessed by daily palpation until day 100, and rejection was defined by the cessation of palpable contractions. Treatment was done as previously described including day 6. All results were analyzed by the Manntel & Haenszel procedure for the nonparametric Log Rank test, validated by the Statistical Analysis System (SAS, Cary, NC).

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